



Oregon State
University

IPM Strategies for Vegetable Crops

Surendra Dara PhD, DAIT

Director, North Willamette Research and Extension Center

Professor, Department of Horticulture

Oregon State University

Surendra.Dara@oregonstate.edu

Pacific Northwest Vegetable Association Conference 16 November 2023

 @calstrawberries @calveggies

 @surendra.dara

 @surendradara

eJournals: ucanr.edu/JEB



and ucanr.edu/pestnews



Common pests of vegetables

Coleoptera

- Click beetle
- Colorado potato beetle
- Flea beetle
- Japanese beetle
- Pepper weevil
- Spotted cucumber beetle
- Striped cucumber beetle
- Wireworms

Dermaptera

- Earwig

Diptera

- Cabbage maggot
- Carrot rust fly
- Leafminers
- Seedcorn maggot

Hemiptera

- Aphids
- Bagrada bug
- Brown marmorated stink bug
- False chinch bug
- Leafhopper
- Lygus bug
- Psyllids
- Squash bug
- Tomato bug
- Whiteflies

Lepidoptera

- Armyworms
- Budworms
- Cutworms
- Diamondback moth
- Earworms
- Fruitworms
- Hornworms
- Imported cabbageworm
- Leafrollers
- Loopers
- Potato tuberworm
- Tomato pinworm

Orthoptera

- Grasshopper

Acarina

- Bulb mite
- Spider mites

Thysanoptera

- Thrips

Others

- Symphylans
- Spotted snake millipedes
- Slugs

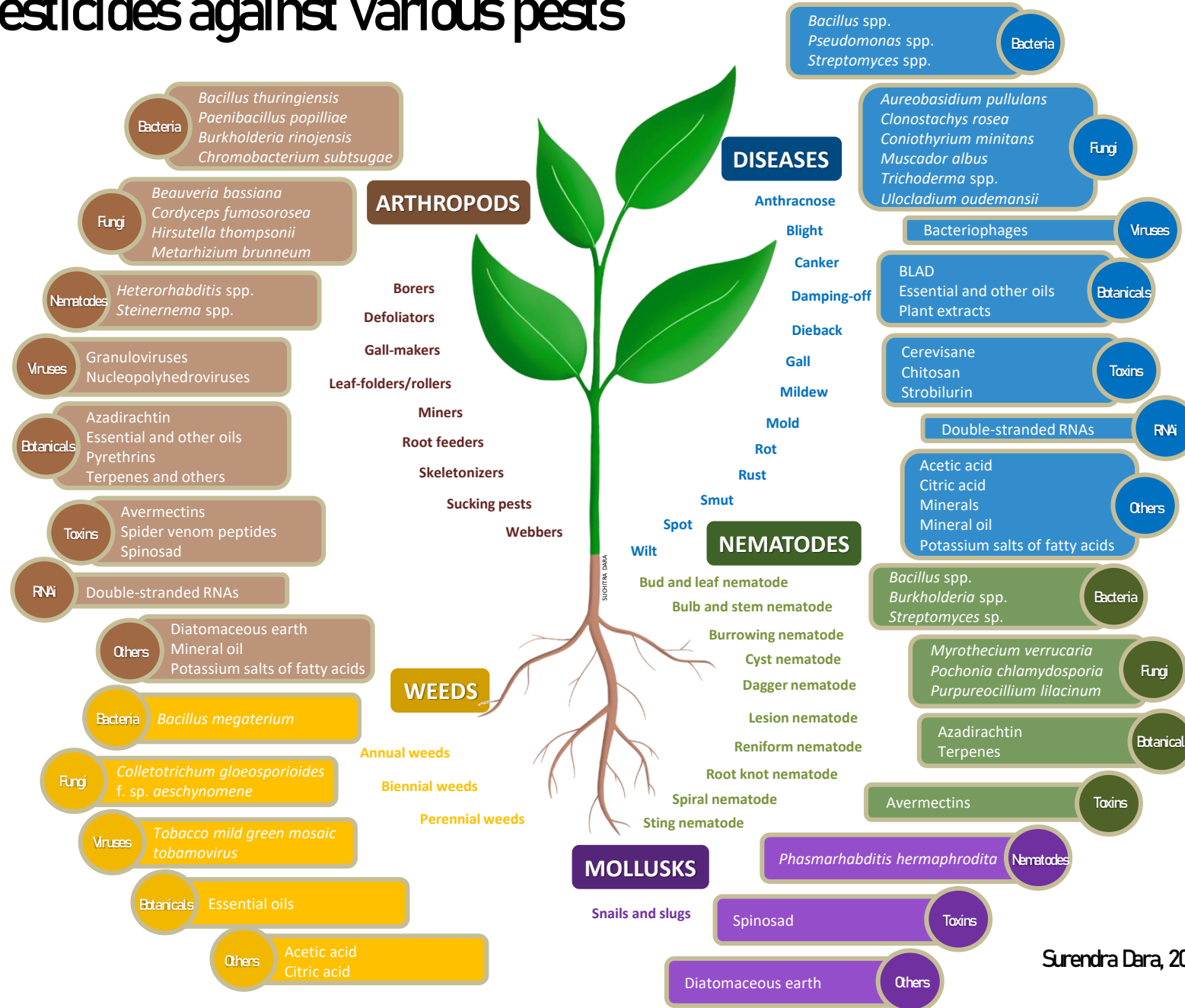
Feeding habit: Chewing, boring, rasping/scraping, piercing and sucking, etc.

Feeding habitat: Surfaces or bored plant tissues (leaves, roots, stems, or fruits), mines, rolls, folds, etc.

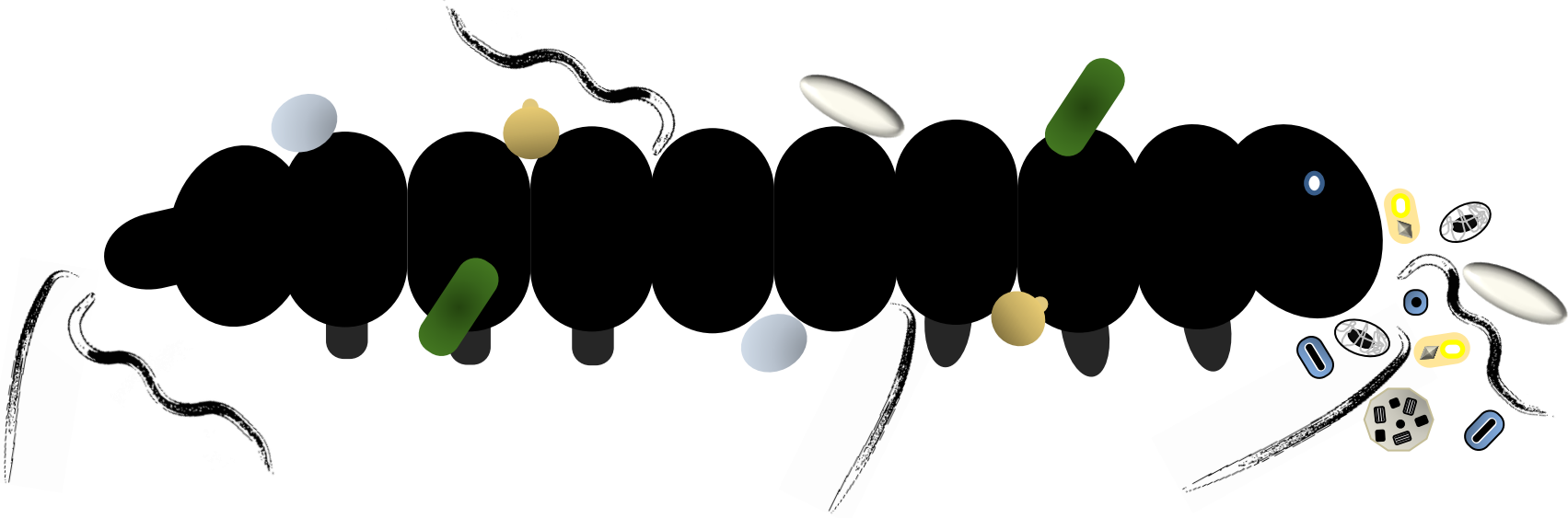
Control option: Based on pest biology, feeding behavior/habitat, mode of action of the control option, prevention/cure, environmental conditions, and others

Biopesticides

Biopesticides against various pests

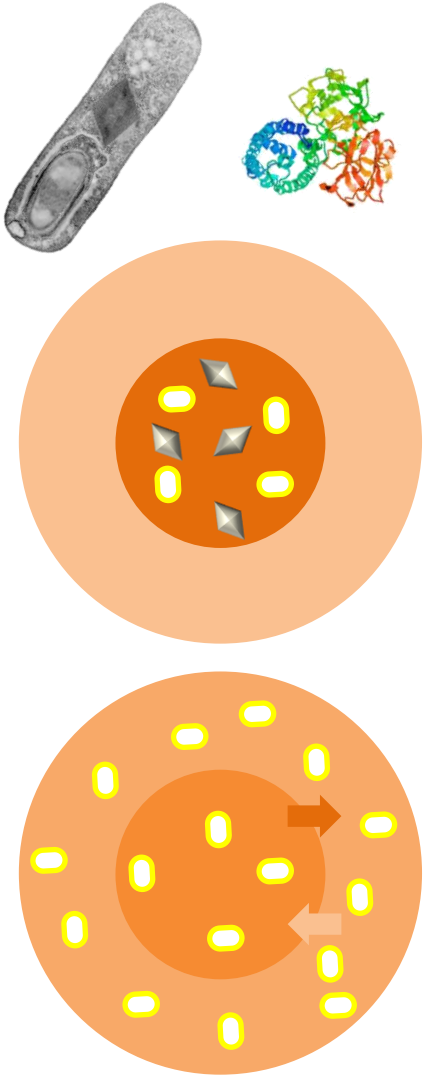


Entomopathogens



Mechanisms of infection

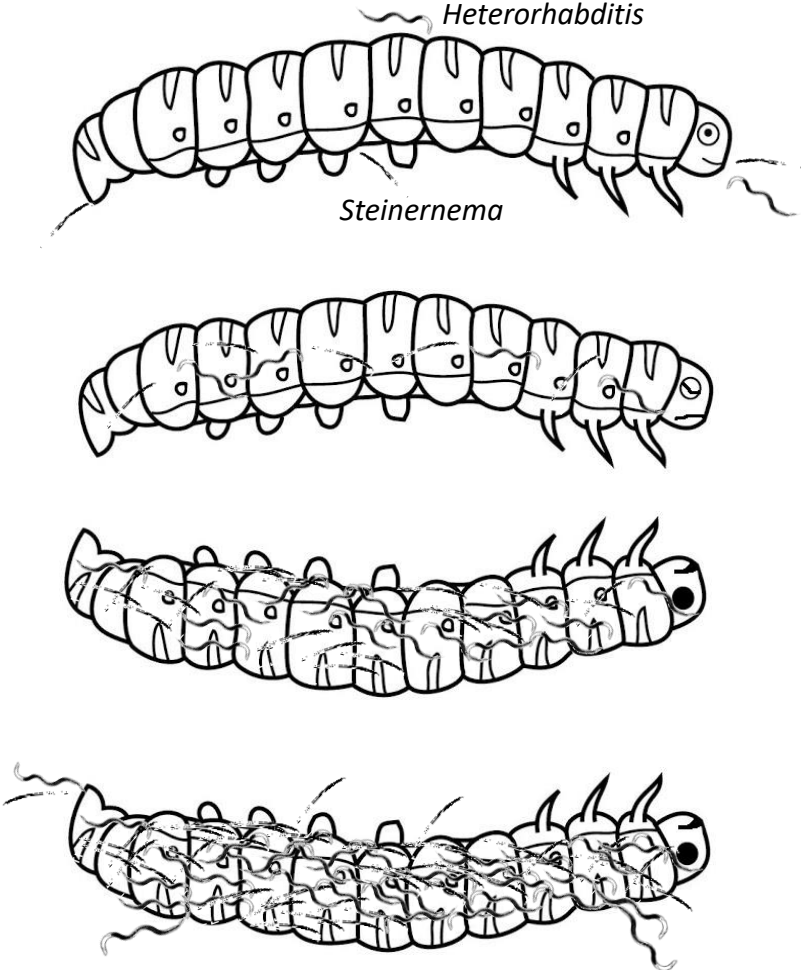
Bacillus thuringiensis



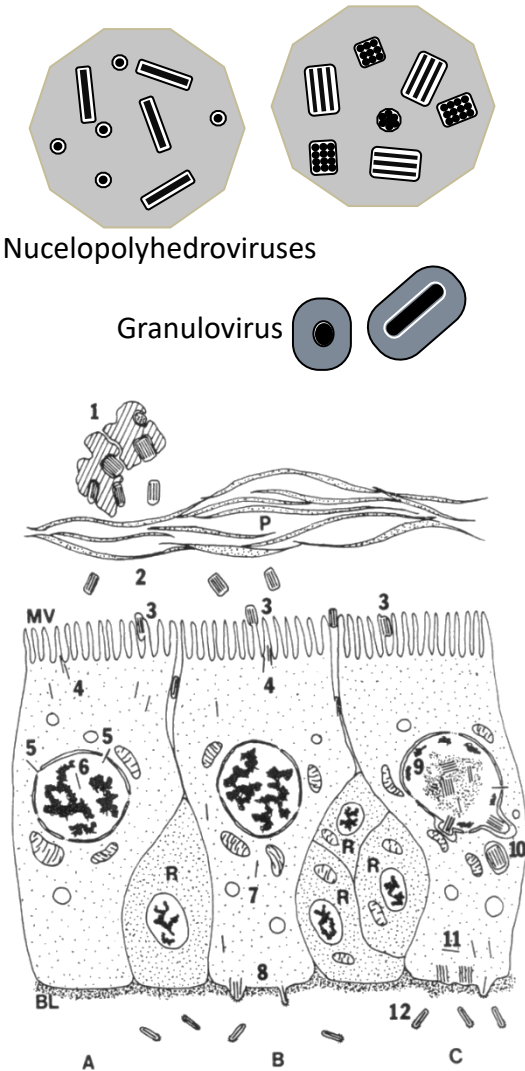
Entomopathogenic Fungi



Entomopathogenic Nematodes



Baculoviruses



Modes of action

| AI | IRAC | Mode of Action |
|-----------------------------------------------------------------------------------------------|------|------------------------------------------------------------------------------------------------------------------|
| Pyrethrins <i>(Chrysanthemum cinerariaefolium)</i> | 3 | Sodium channel modulators causing hyperexcitation and sometimes nerve block |
| Spinosyns <i>(Polysaccharospora spinosa)</i> | 5 | Nicotinic acetylcholine receptor allosteric modulators – Site I causing hyperexcitation of the nervous system |
| Avermectins <i>(Streptomyces avermitilis)</i> | 6 | Glutamate-gated chloride channel allosteric modulators causing paralysis |
| { Spider venom peptide (GS-omega/kappa HTX-Hv1a peptide) (Australian funnel-web spider) | 32 | Nicotinic acetylcholine receptor allosteric modulators – Site II causing hyperexcitation } of the nervous system |
| Azadirachtin <i>(Azadirachta indica)</i> | UN | Unknown/uncertain Insecticide/antifeedant/repellent/IGR |
| Botanical extracts/oils | UNE | Unknown/uncertain Insecticide/antifeedant/repellent |

Modes of action

AI

Diatomaceous earth

Mineral oil

RNAi

IRAC

UNM

UNM

(35)

Mode of Action

Cuticle abrasion and absorption of lipids

Asphyxiation

Targeted protein suppression



Insecticide Resistance Action Committee

<https://irac-online.org/documents/moa-classification/>

IRAC

Insecticide Resistance Action Committee
Mode of Action Classification

Key to Targeted Physiology

- Nerve & Muscle
- Growth & Development
- Respiration
- Midgut
- Unknown or Non-specific

Group 1: Acetylcholinesterase (AChE) inhibitors
(Only representative actives of the groups are shown)

1A Carbamates: Carbaryl, Carbosulfan, Methomyl

1B Organophosphates: Acephate, Chlorpyrifos, Phorate

Group 2: GABA-gated chloride channel antagonists

2A Cyclopyridone Organochlorines: Chlorfens, Endosulfan

2B Phenylpyrazoles (Fiproles): Ethiprole, Fipronil

Group 3: Sodium channel modulators (Only representative actives of group 3A are shown)

3A Pyrethroids Pyrethrins: Bifenthrin, Deltamethrin, Etofenprox, Permethrin, Lambda-cyhalothrin, Taluthrin

3B DDT, Methoxychlor

Group 4: Nicotinic acetylcholine receptor (nAChR) competitive modulators

4A Neonicotinoids: Clothianidin, Imidacloprid, Thiacloprid

4B Nicotine

4C Sulfoximines: Sulfoxaflor

4D Butenolides: Flupyradfurone

4E Mesoionics: Trifluromethoxyvinyl, Dieldromethozal

4F Pyridylidenes: Flupyrimin

Group 5: Nicotinic acetylcholine receptor (nAChR) allosteric modulators site I

5 Spinosyns: Spinetoram, Spinosad

Group 6: Glutamate-gated chloride channel (GluCl) allosteric modulators

6 Avermectins & Milbemycins: Emamectin benzoate R1, Ivermectin, Milbemectin

Group 7: Juvenile hormone receptor modulators

7A Juvenile hormone analogues: Hydroprone R1 = ethyl, R2 = H; Methoprene R1 = isopropyl, R2 = -OCH₃; Iniproprone R1 = propargyl, R2 = H

7B Fenoxycarb

7C Fenoxycarb

7C Pynproxifen

7C Pyriproxyfen

Group 8: Miscellaneous non-specific (multi-site) inhibitors

8A Alkyl halides: H₃C-Br, Methyl bromide

8B Chloroclorin

8C Fluorides: Na₂BO₇·10H₂O, Borax, BD Borates

8E Tartar emetic

8F Methyl isothiocyanate generators: Dazomet, Metam

Use of Groups:

- Alternations, sequences or rotations of compounds between MoA groups reduce selection for target site resistance.
- Applications are arranged into MoA spray windows defined by crop growth stage and pest biology. Several sprays of a compound may be possible within each spray window, but successive generations of a pest should not be treated with compounds from the same MoA group. Local expert advice on spray windows and timings should always be followed.
- Groups in the classification whose members do not act at a common target site are exempt from the prescription against rotation within the group (Groups 9, 13 and all UN groups; UN, UNE, UNF, UNM, UNP & UNV).

Use of Sub-Groups:

- Sub-groups represent distinct structural classes which are believed to have the same mode of action.
- Sub-groups provide differentiation between compounds that may bind at the same target site but are structurally different enough that risk of metabolic cross-resistance is lower than for close chemical analogs.
- Cross-resistance potential between sub-groups is higher than between groups, so rotation between sub-groups should be considered only when there are no alternatives, and only if cross-resistance does not exist, following consultation with local expert advice. These exceptions are not sustainable, and alternative options should be sought.

Group 9: Chordotonal organ TRPV channel modulators

9B Pyridine azomethine derivatives: Pymetrozine, Pyrifluquinazon

9D Pyropenes: Abdoxypropan

Group 10: Mile growth inhibitors affecting CHS1

10A: Clofentazine, Clofentezine, Heptythiazox

10B: Etoxazole

Group 11: Microbial disruptors of insect midgut membranes

Includes transgenic crops expressing *Bacillus thuringiensis* toxins (however, specific guidance for resistance management of transgenic crops is not based on rotation of modes of action)

Rotation between certain specific B.t. microbial products may provide resistance management benefits for some pests. Consult product-specific recommendations.

11A *Bacillus thuringiensis*

11B *Bacillus sphaericus*

Group 12: Inhibitors of mitochondrial ATP synthase

12A: Diafenthiuron, Diafenthiuron

12B: Organotin miticides: Azoxystrobin, Fenbutathion oxide, Cyhexatin

12C Propargite

12D Tetradifon

Group 13: Uncouplers of oxidative phosphorylation via disruption of proton gradient

13 Pyridoles, Dinitrophenols, Sulfuramid

DNOC

Group 14: Nicotinic acetylcholine receptor (nAChR) channel blockers

14 Nereistoxin analogues: Benalipax, Carbap hydrochloride, Thiocyanam, Thiocyanamide sodium

Group 15: Inhibitors of chitin biosynthesis affecting CHS1
(Only representative actives of group are shown)

Diflubenzuron

Flufenoxuron

Lufenuron

Novaluron

Teflubenzuron

15 Benzoylureas

Group 16: Inhibitors of chitin biosynthesis, type 1

Buprofezin

16 Buprofezin

Group 18: Ecdysone receptor agonists

Chromafenozide

Halofenozide

Methoxyfenozide

Tebufenozide

18 Diacylhydrazines

Group 19: Octopamine receptor agonists

Amirbaz

19 Amirbaz

Group 20: Mitochondrial complex III electron transport inhibitors – Qo site

20A Hydranmethylin

20B Acequinolyf

20C Fluacrypyrim

20D Bifenazate

Disclaimer: While CropLife International and IRAC make every effort to present accurate and reliable information, they do not guarantee the accuracy, completeness, efficacy, timeliness, or correct sequencing of such information. Inclusion of active ingredients in the IRAC Code Lists is based on scientific evaluation of their mode of action; it does not provide any kind of testimonial for the use of a product or a judgment of efficacy. CropLife International and IRAC are not responsible for, and expressly disclaim all liability for, damages of any kind arising out of use, reference to, or reliance on information provided. Listing of chemical classes or modes of action must not be interpreted as approval for use of a compound in a given country. Prior to implementation, each user must determine the current registration status in the country of use and strictly adhere to the uses and instructions approved in that country.

Group 21: Mitochondrial complex I electron transport inhibitors

Fenazaquin

Pyridaben

Fenpyroximate

Pyrimifan

Tolfenpyrad

Tebufenpyrad

21B Rotenone

21A METI acaricides and insecticides

Group 22: Voltage-dependent sodium channel blockers

Indoxacarb

22A Oxadiazines

22B Semicarbazones

Metarfluzone

Group 23: Inhibitors of acetyl CoA carboxylase

Spirodifen

Spiromesifen

Spirotetramat

23 Tetrone & Tetramic acid derivatives

Group 24: Mitochondrial complex IV electron transport inhibitors

AIP

PHs

CasPz

24A Phosphides

ZnSPz

Zinc phosphide

CN-

Cyanide salts

24B Cyanides

Group 25: Mitochondrial complex II electron transport inhibitors

Cyanothiazole

Cyflumetofen

25A beta-Ketonitrile derivatives

25B Carboxanilides

Group 26: Ryanodine receptor modulators

Chlorantraniliprole R=Cl

Cyrantraniliprole R=CN

Flubendiamide

Cyflumetofen

Tetrafluprole

26 Diamides

Group 29: Chordotonal organ neotamidaase inhibitors

Fonitamid

29 Fonitamid

Group 30: GABA-gated chloride channel allosteric modulators

Brotianilide

Fluxametamide

Isoxazolines

30 Mela-diamides & Isoxazolines

Group 31: Baculoviruses

Cydia pomonella GV

Thaumetobia lacticola GV

Articaria gemmatata MNPV

Heliothis virescens NPV

31 Granuloviruses & Nucleopolyhedroviruses

Group 32: Nicotinic Acetylcholine receptor (nAChR) allosteric modulators site II

GS-omega/kappa HXTX-Hv1a peptide

32 GS-omega/kappa HXTX-Hv1a peptide

Group 33: Calcium-activated potassium channel (KCa2) modulators

Acynonapyr

33 Acynonapyr

Group 34: Mitochondrial complex III electron transport inhibitors – Q1 site

Flometoquin

34 Flometoquin

Group 36: Chordotonal organ modulators – undesignated target site

Dimpropridaz

36 Dimpropridaz

UN: Unknown or uncertain mode of action

Beauveria bassiana strains

Metarhizium brunneum strain F52

Pleurochaete funiculosus

Apoka strain 9F

Burkholderia spp.

Wobaschia gigavita (Zap)

UNB Bacterial agents (non-Bt)

UNM Non-specific mechanical and physical disruptors

Diatomaceous earth

Mineral oil

UNE Botanical essence including synthetic, extracts and unrefined oils

Chenopodium ambrosioides near ambrosioides extract

Fatty acid monomers with glycerol or propanediol

Neem oil

Poster Notes:

- Sub-group 3B: DDT is no longer used in agriculture and therefore this is only applicable for the control of insect vectors of human disease, such as mosquitoes, because of a lack of alternatives.
- Sub-group 10A: Heptythiazox is grouped with Clofentezine because they exhibit cross-resistance even though they are structurally distinct. Difenoxazin has been added to this group because it is a close analogue of Clofentezine and is expected to have the same mode of action.
- Group 20: While there is strong evidence that Bifenazate acts on the Qo site of Mitochondrial Complex III and some Bifenazate resistance mutations confer cross-resistance to Acequinolyf, the sites of action of Fluacrypyrim and Hydranmethylin have not been determined.
- Groups 26, 27 and 35 are unassigned.
- In some cases, only representative actives are shown.
- Please visit www.irac-online.org for the complete IRAC classification.



CropLife

New IPM Model



Effective management strategies

- Integrated pest management
 - Take advantage of all available options
 - Consider additive and synergistic effect of multiple control options
 - Combine and rotate synthetic and biological pesticides
 - Follow IPM principles beyond pest management
- When using biopesticides, understand their mode of action, compatibility, storage and handling, and other use strategies
- IPM = GAP (Good Agricultural Practices) = CCC (Comprehensive Crop Care)
- Insecticide resistance management
 - Pests develop resistance to all kinds of pesticides
 - Avoid repetitive use of the same control option that has a risk of resistance development
- Balanced approach for both short-term and long-term benefits
- Use of modern technologies that enhance pest control efficacy

BURLEIGH DODDS SERIES IN AGRICULTURAL SCIENCE

Improving integrated pest management in horticulture

Edited by Professor Rosemary Collier, Warwick University, UK



bd burleigh dodds
SCIENCE PUBLISHING

BURLEIGH DODDS SERIES IN AGRICULTURAL SCIENCE

Advances in biostimulants as an integrated pest management tool in horticulture

Surendra K. Dara, University of California Cooperative Extension, USA



bd burleigh dodds
SCIENCE PUBLISHING





Thank you!



Download free **IPMinfo** app



<https://tinyurl.com/eJournalEB>
<https://tinyurl.com/eJournalPestNews>



@calstrawberries and @calveggies



@surendradara



@surendra.dara



<https://youtube.com/c/SurendraDara>



Surendra.Dara@oregonstate.edu

